# GUIDELINES FOR SAMPLING FRESHWATER MUSSELS IN INDIANA



Wildlife Section
Division of Fish & Wildlife
Indiana Department of Natural Resources

**Third Edition (December 2014)** 

The following guidelines are intended to provide accuracy and consistency for the mussel surveys completed in the waters of Indiana for the purposes of environmental review, while protecting the state's mussel resource.

## General guidelines that apply to all mussel surveys:

- The **impact area** is defined as the entire area where in-channel work will occur.
- The **study area** includes the impact area, as well as an area upstream and downstream from the impact area boundaries. These distances are determined by multiplying the average stream width for the impact area by 2 (for upstream) and by 4 (for downstream); a minimum of 50 meters upstream and 100 meters downstream from the impact area boundaries is required for all surveys.
- Those persons conducting mussel surveys in Indiana must have an adequate knowledge of Midwestern mussel communities and be able to correctly identify live individuals of Indiana's mussel fauna (Table 1).
- Those persons conducting mussel surveys must be able to obtain an Indiana Scientific Purposes License (contact Operations Section Staff Specialist at: <a href="mailto:lpetercheff@dnr.IN.gov">lpetercheff@dnr.IN.gov</a> or 317-233-6527) as well as a Federal Fish and Wildlife Permit (if required for survey location).
- Water flows must be at normal levels before mussel surveys can be completed. Surveys can only be completed when discharge for the closest USGS gage (<a href="http://waterwatch.usgs.gov/new/index.php?m=real&r=in&w=map">http://waterwatch.usgs.gov/new/index.php?m=real&r=in&w=map</a>) is at or below its median daily streamflow value for the dates of the survey. Depending on the time of year, certain surveys may not be dependent on this criteria; requests to sample at higher discharges should be forwarded to the Nongame Aquatic Biologist (contact information below).
- Surveys can only be completed between May 1-October 15; these restraints could be modified for unusually high spring/fall water temperatures. Surveys can not be conducted when air temperatures are at or below 32°F.
- All live mussels collected during surveys must be identified to species, enumerated and measured (height and length in millimeters).
- Proper handling of live mussels (including keeping mussels in flow-through river bags etc.) must be utilized to minimize negative impacts during handling. All live mussels collected during survey efforts must be returned to the study area from which they came. When water temperatures are above 50°F, live mussels can be simply placed back on the substrate surface; when water temperatures are below 50°F, live mussels must be replanted into the substrate by hand.
- At least one valve for each species collected during surveying must be vouchered for verification purposes. No live mussels are to be sacrificed for this purpose. For species only found live within the study area, high-quality digital images of both valves and beak should be substituted.
- Vouchered shell material and digital images must be provided to the Nongame Aquatic Biologist for verification. Shell material of significance will be deposited in the Indiana State Museum.

- Upon completion of the mussel survey, a final report must be prepared that includes all of the following:
  - o a complete list of all mussel species (including exotics) collected (live or dead) during survey activities within the study area with indication of best condition of shell material (live, fresh dead, weathered dead or subfossil) encountered
  - o measurement information for all live mussels
  - o a detailed description of the methods and effort used to complete the survey (as defined by these guidelines)
  - o the exact location (using GPS coordinates) of any live federal/state endangered mussels (Table 1) collected during survey activities.
  - o a detailed map of the study and impact areas, delineating substrate types and water depths throughout the study area
  - o information on the water temperature, air temperature and water clarity for the dates of surveying
  - o pictures clearly identifying the impact area and study area
- A copy of the final report must be provided to the following Indiana Department of Natural Resources staff: 1) Operations Staff Specialist (as required by the Scientific Purposes License), 2) Environmental Biologist requesting the survey, and 3) Nongame Aquatic Biologist.

## **Guidelines specific for Nonwadeable and Wadeable stream reaches:**

#### NONWADEABLE:

- **Nonwadeable** reaches are defined as those where greater than 25% of the bottom of the study area cannot be adequately sampled without the use of diving methods.
- The following reaches are considered nonwadeable regardless of reach characteristics:
  - 1) entire Ohio River
  - 2) St. Joseph River in St. Joseph and Elkhart counties
  - 3) Maumee River in Allen County
  - 4) Wabash River from the Ohio River upstream to Huntington Reservoir
  - 5) White River upstream to the confluence of the East Fork and West Fork
  - 6) West Fork White River from confluence with White River upstream to the Johnson/Morgan county line
  - 7) East Fork White River from confluence with the White River upstream to Columbus (Bartholomew County)
- All nonwadeable reaches will require surveys to be completed by diving.
- Quantitative surveys using transects perpendicular to stream flow will be required.
- The distance between transects will be determined by dividing the average stream width within the impact area by two. The maximum distance between transects will be 100 meters.
- The number of transects will be determined by dividing the distance between transects into the total length of the study area, with at least one transect encompassing each of the upstream and downstream boundaries of the study area.
- Along each transect, at 10 evenly-spaced increments, mussels will be collected from 2-1/4 m<sup>2</sup> quadrats. All substrate to a depth of 15-20 cm must be excavated, bagged and processed stream-side.

- All transect endpoints and location of quadrats along the transects must be identified using GPS coordinates. A detailed map depicting the study area boundaries and location of transects/quadrats must be included in the final report.
- A detailed map depicting the mussel density per quadrat within the study area must be included in the final report.
- More intensive surveying within the areas of highest mussel densities within the study area determined by the initial transect sampling may be required in reaches that possibly harbor federal/state endangered mussel species.
- For all nonwadeable sites, additional qualitative sampling must be conducted in shallow areas, if present, within the project area. This would include areas around exposed sand/gravel bars, backwaters, side-channels, etc. These should be sampled using techniques described under the 'wadeable' portion of these protocols.
- Ohio River sampling may require special sampling protocols currently being developed by the Ohio River Valley Ecosystem Mollusk Subgroup. The Nongame Aquatic Biologist should be contacted for the most current version of these protocols (see below for contact information).

### **WADEABLE:**

- Wadeable reaches are defined as those where greater than 75% of the bottom of the entire study area can be adequately sampled without the use of diving methods.
- Qualitative sampling methods are sufficient for wadeable stream reaches.
- The entire study area, including all habitats, must be visually (if possible) and physically searched for live mussels and dead shell material.
- Physical search techniques must be attempted in order to collect small individuals, small species and species that tend to bury deeper in the substrate. This could include searching through the substrate with hands, shoed-feet or gently pulled specialized rakes.
- All exposed areas and stream banks within the study area must be searched for dead shell material.
- Person-hours spent sampling must be included in the final report.
- Even though transect sampling is not required during surveys of wadeable reaches, maps depicting the areas of highest mussel concentrations and describing the substrate types throughout the study area must be included in the final report.

#### **Guidelines prepared by:**

Brant E. Fisher, Nongame Aquatic Biologist Atterbury Fish & Wildlife Area 7970 South Rowe Street PO Box 3000 Edinburgh, IN 46124-3000

email: bfisher@dnr.IN.gov phone: 812-526-5816

TABLE 1: NATIVE FRESHWATER MUSSELS OF INDIANA:

IAD	LE I. NATIVE	FRESHWATER MUS	SELS OF INDIANA:	
	GENUS	SPECIES	COMMON NAME	STATUS
1	Actinonaias	ligamentina	MUCKET	
2	Alasmidonta	marginata	ELKTOE	
3	Alasmidonta	viridis	SLIPPERSHELL MUSSEL	
4	Amblema	plicata	THREERIDGE	
5	Anodonta	suborbiculata	FLAT FLOATER	
6	Anodontoides	ferussacianus	CYLINDRICAL PAPERSHELL	
7	Arcidens	confragosus	ROCK POCKETBOOK	
8	Cumberlandia	monodonta	SPECTACLECASE	extirpated
9	Cyclonaias	tuberculata	PURPLE WARTYBACK	
10	Cyprogenia	stegaria	FANSHELL	federal endangered
11	Ellipsaria	lineolata	BUTTERFLY	
12	Elliptio	crassidens	ELEPHANTEAR	
13	Elliptio	dilatata	SPIKE	
14	Epioblasma	flexuosa	LEAFSHELL	extirpated
15	Epioblasma	obliquata obliquata	CATSPAW	extirpated
16	Epioblasma	obliquata perobliqua	WHITE CATSPAW	federal endangered
17	Epioblasma	personata	ROUND COMBSHELL	extirpated
18	Epioblasma	propinqua	TENNESSEE RIFFLESHELL	extirpated
19	Epioblasma	sampsonii	WABASH RIFFLESHELL	extirpated
20	Epioblasma	torulosa rangiana	NORTHERN RIFFLESHELL	federal endangered
21	Epioblasma	torulosa torulosa	TUBERCLED BLOSSOM	federal endangered
22	Epioblasma	triquetra	SNUFFBOX	federal endangered
23	Fusconaia	ebena	EBONYSHELL	
24	Fusconaia	flava	WABASH PIGTOE	
25	Fusconaia	subrotunda	LONGSOLID	state endangered
26	Hemistena	lata	CRACKING PEARLYMUSSEL	extirpated
27	Lampsilis	abrupta	PINK MUCKET	federal endangered
28	Lampsilis	cardium	PLAIN POCKETBOOK	
29	Lampsilis	fasciola	WAVYRAYED LAMPMUSSEL	special concern
30	Lampsilis	ovata	POCKETBOOK	
31	Lampsilis	siliquoidea	FATMUCKET	
32	Lampsilis	teres	YELLOW SANDSHELL	
33	Lasmigona	complanata	WHITE HEELSPLITTER	
34	Lasmigona	compressa	CREEK HEELSPLITTER	
35	Lasmigona	costata	FLUTEDSHELL	
36	Leptodea	fragilis	FRAGILE PAPERSHELL	
37	Leptodea	leptodon	SCALESHELL	extirpated
38	Ligumia	recta	BLACK SANDSHELL	
39	Ligumia	subrostrata	PONDMUSSEL	
40	Megalonaias	nervosa	WASHBOARD	
41	Obliquaria	reflexa	THREEHORN WARTYBACK	
42	Obovaria	olivaria	HICKORYNUT	
43	Obovaria	retusa	RING PINK	extirpated
44	Obovaria	subrotunda	ROUND HICKORYNUT	state endangered

45	Plethobasus	cicatricosus	WHITE WARTYBACK	federal endangered
46	Plethobasus	cooperianus	ORANGEFOOT PIMPLEBACK	federal endangered
47	Plethobasus	cyphyus	SHEEPNOSE	federal endangered
48	Pleurobema	clava	CLUBSHELL	federal endangered
49	Pleurobema	cordatum	OHIO PIGTOE	special concern
50	Pleurobema	plenum	ROUGH PIGTOE	federal endangered
51	Pleurobema	rubrum	PYRAMID PIGTOE	state endangered
52	Pleurobema	sintoxia	ROUND PIGTOE	
53	Potamilus	alatus	PINK HEELSPLITTER	
54	Potamilus	сарах	FAT POCKETBOOK	federal endangered
55	Potamilus	ohiensis	PINK PAPERSHELL	
56	Ptychobranchus	fasciolaris	KIDNEYSHELL	special concern
57	Pyganodon	grandis	GIANT FLOATER	
58	Quadrula	cylindrica cylindrica	RABBITSFOOT	state end/fed threatened
59	Quadrula	fragosa	WINGED MAPLELEAF	extirpated
60	Quadrula	metanevra	MONKEYFACE	
61	Quadrula	nodulata	WARTYBACK	
62	Quadrula	pustulosa pustulosa	PIMPLEBACK	
63	Quadrula	quadrula	MAPLELEAF	
64	Simpsonaias	ambigua	SALAMANDER MUSSEL	special concern
65	Strophitus	undulatus	CREEPER	
66	Toxolasma	lividum	PURPLE LILLIPUT	special concern
67	Toxolasma	parvum	LILLIPUT	
68	Toxolasma	texasense	TEXAS LILLIPUT	
69	Tritogonia	verrucosa	PISTOLGRIP	
70	Truncilla	donaciformis	FAWNSFOOT	
71	Truncilla	truncata	DEERTOE	
72	Uniomerus	tetralasmus	PONDHORN	
73	Utterbackia	imbecillis	PAPER PONDSHELL	
74	Venustaconcha	ellipsiformis	ELLIPSE	special concern
75	Villosa	fabalis	RAYED BEAN	federal endangered
76	Villosa	iris	RAINBOW	
77	Villosa	lienosa	LITTLE SPECTACLECASE	special concern
				-